



Detecting Infectious Viral Pathogens Toward Developing Smart Water Reuse Disinfection Systems

Benito J. Mariñas

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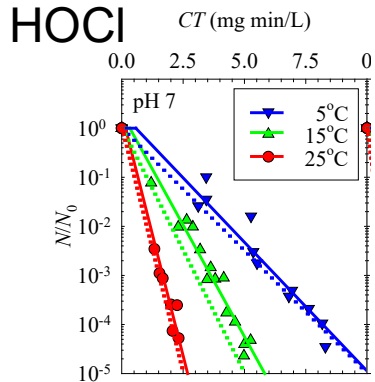
University of Illinois at Urbana-Champaign, USA



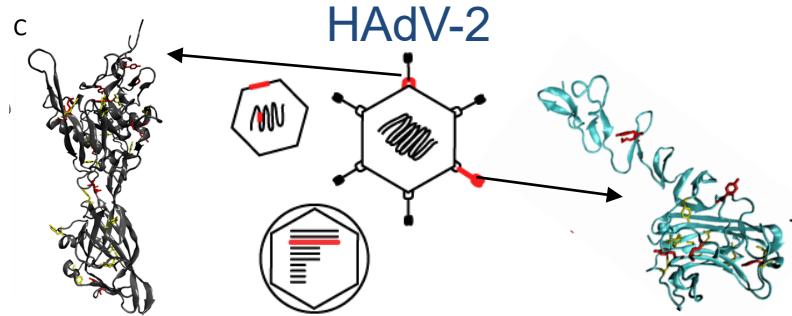
Inter/Transdisciplinary Research Approach

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Inactivation kinetics

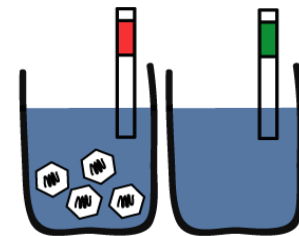
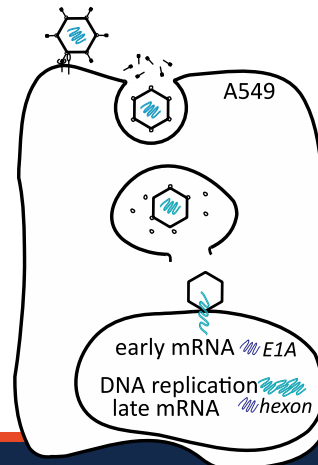
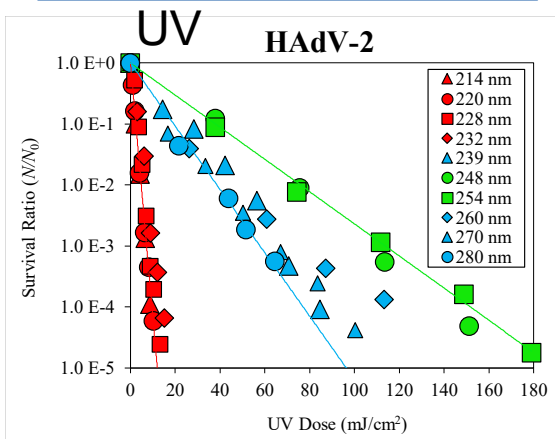


Protein/Genome transformation

Replication cycle disruption



Sensor development



Gall et al., *PLoS Pathog*
2015, 11 (6), e1004867



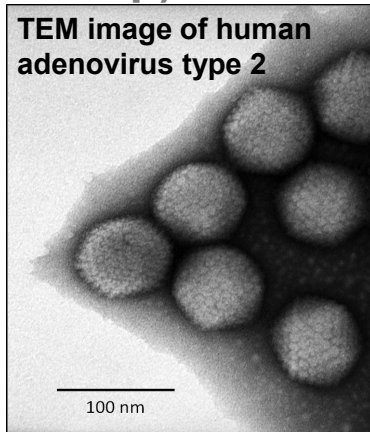
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Viral Pathogens, Disinfectants & Objectives

Martin Page, Kwanrawee Sirikanchana, Aimee Gall, Bernardo Vazquez, Kelley Gonçalves, Wen Cong, Anisa Hardin
Joanna L. Shisler

Pathogens:

- Human Adenovirus HAdV-2 (HAdV species C; non-enveloped capsid ~90 nm; dsDNA ~36 kbp)



- Coxsackievirus B5 (enterovirus species B; non-enveloped capsid ~30 nm; +ssRNA ~7.4 kb)

Disinfectants:

- Free chlorine
- Monochloramine
- UV light
 - Experiments performed with synthetic buffered solutions in batch reactors
 - Viability assessment by plaque assay using human lung A549 carcinoma cells (HAdV-2), or Buffalo Green Monkey Kidney (BGMK) cells (CoxV-B5)

Broad Objective:

- Achieve better control of viruses in water reuse
 - Determination of inactivation kinetics
 - Elucidation of inactivation mechanisms
 - Simple, rapid, robust sensors to **detect infectious viruses** (ultimately including those for which cell culture is not available, e.g., human norovirus)



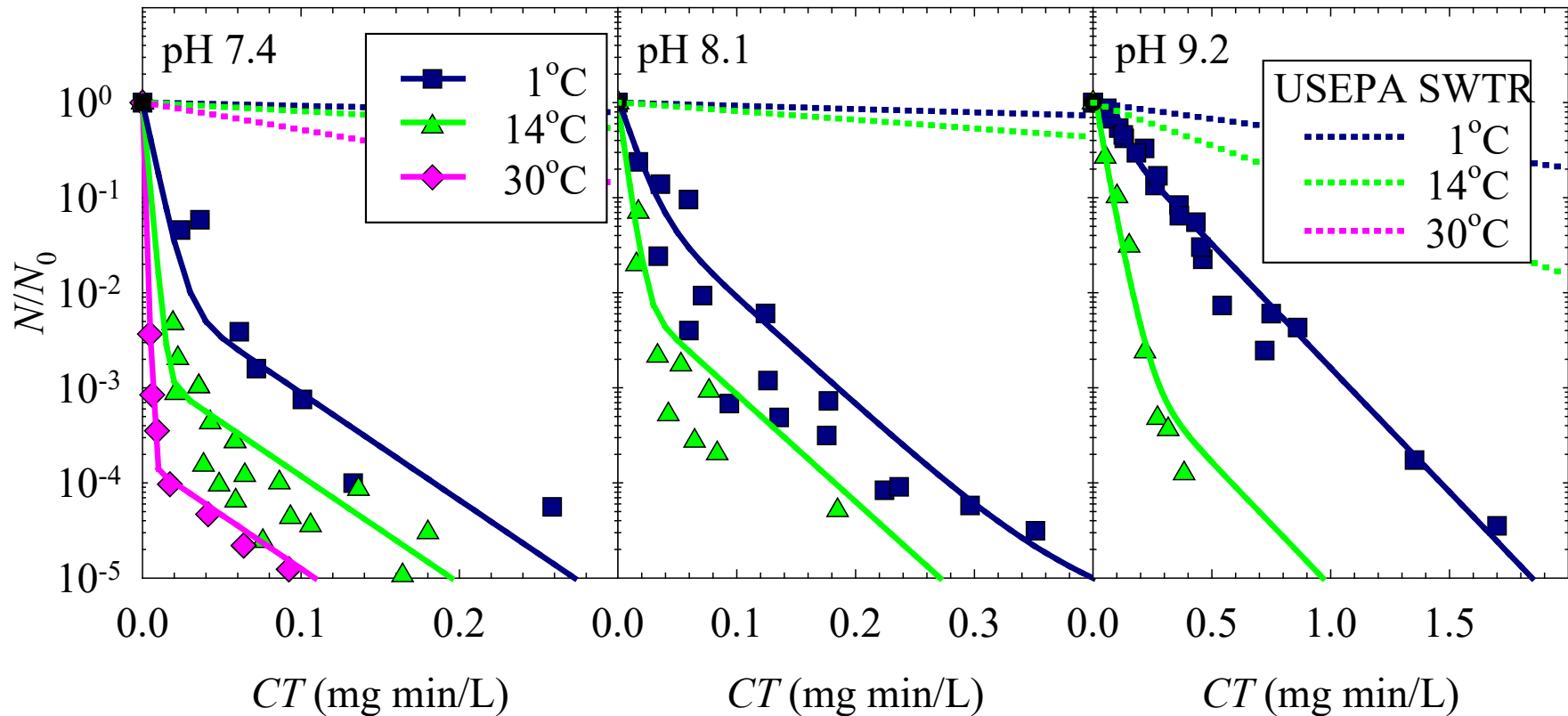


INACTIVATION KINETICS



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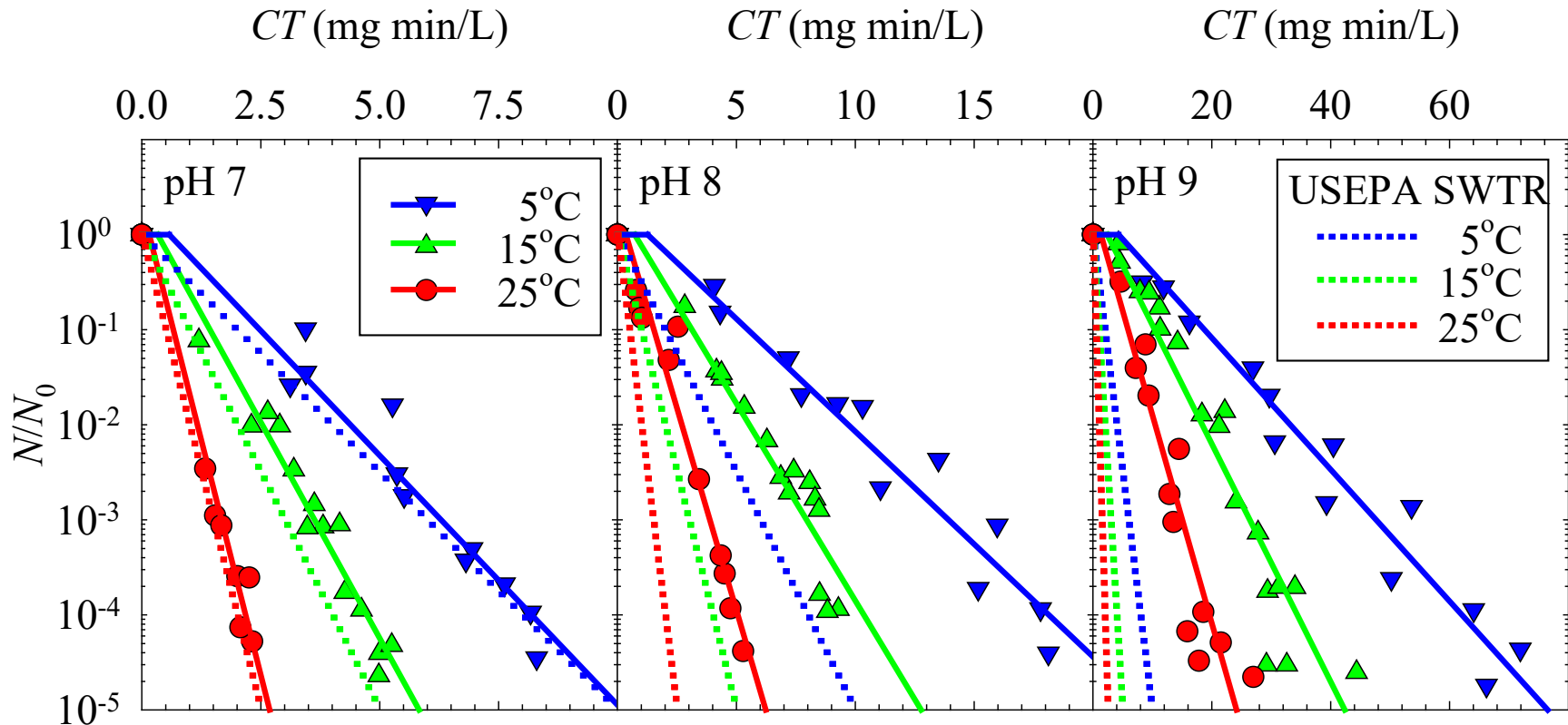
Inactivation of Human Adenovirus 2 with Free Chlorine



M.A. Page, J.L. Shisler, B.J. Marinas, *Wat. Res.*, 2009, 43, 2916-2926



Inactivation of Coxsackievirus B5 with Free Chlorine

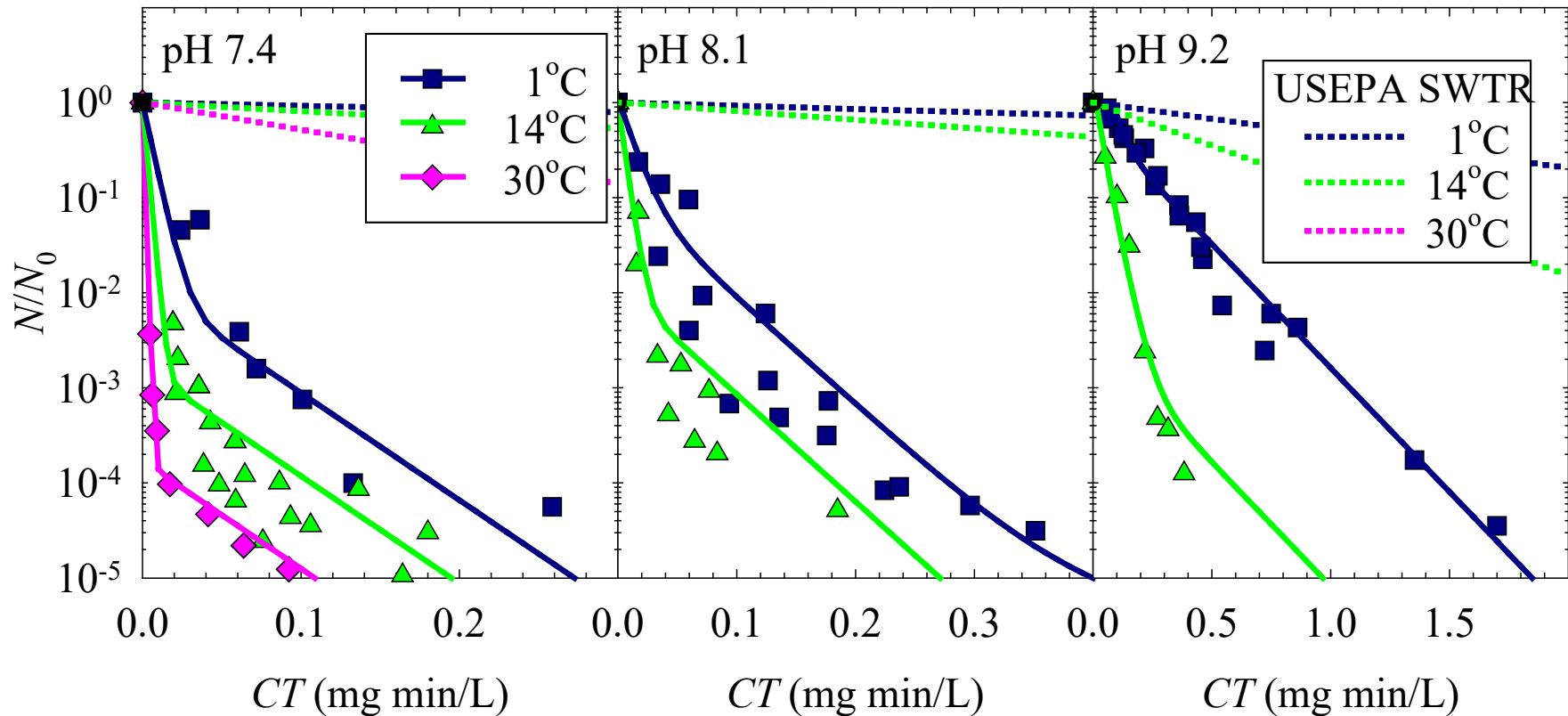


W. Cong, A. Hardin, B.J. Marinas, *in preparation*



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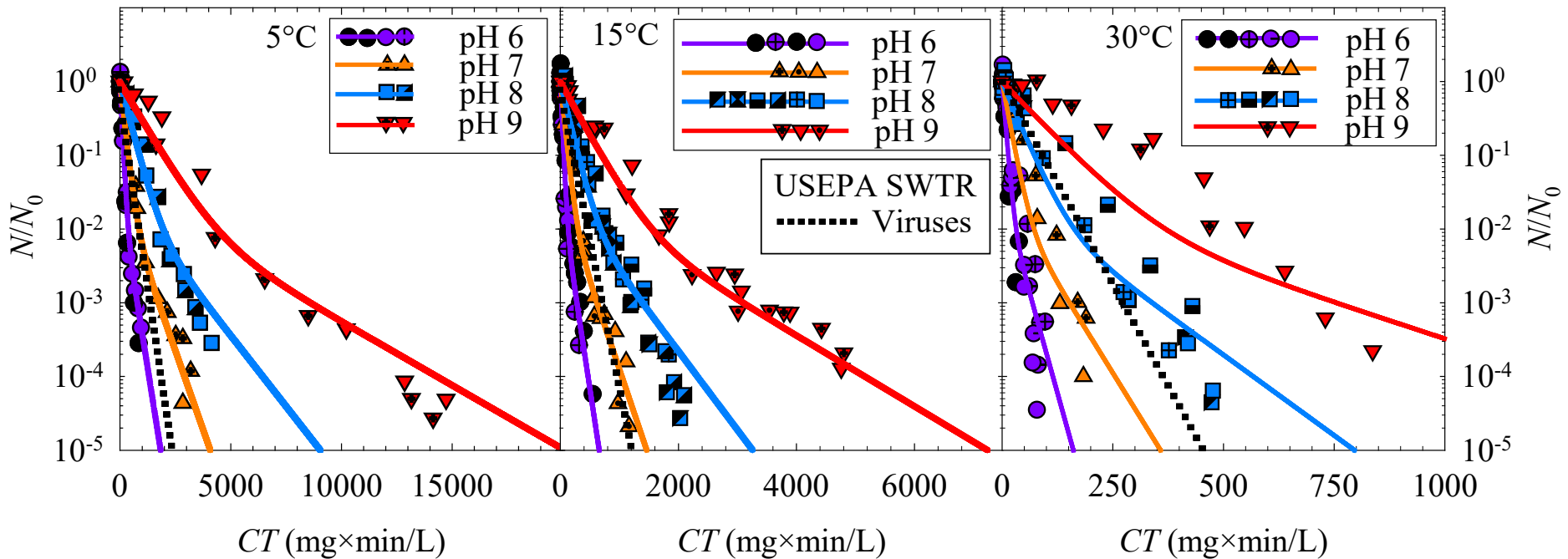
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Inactivation of Human Adenovirus 2 with Monochloramine

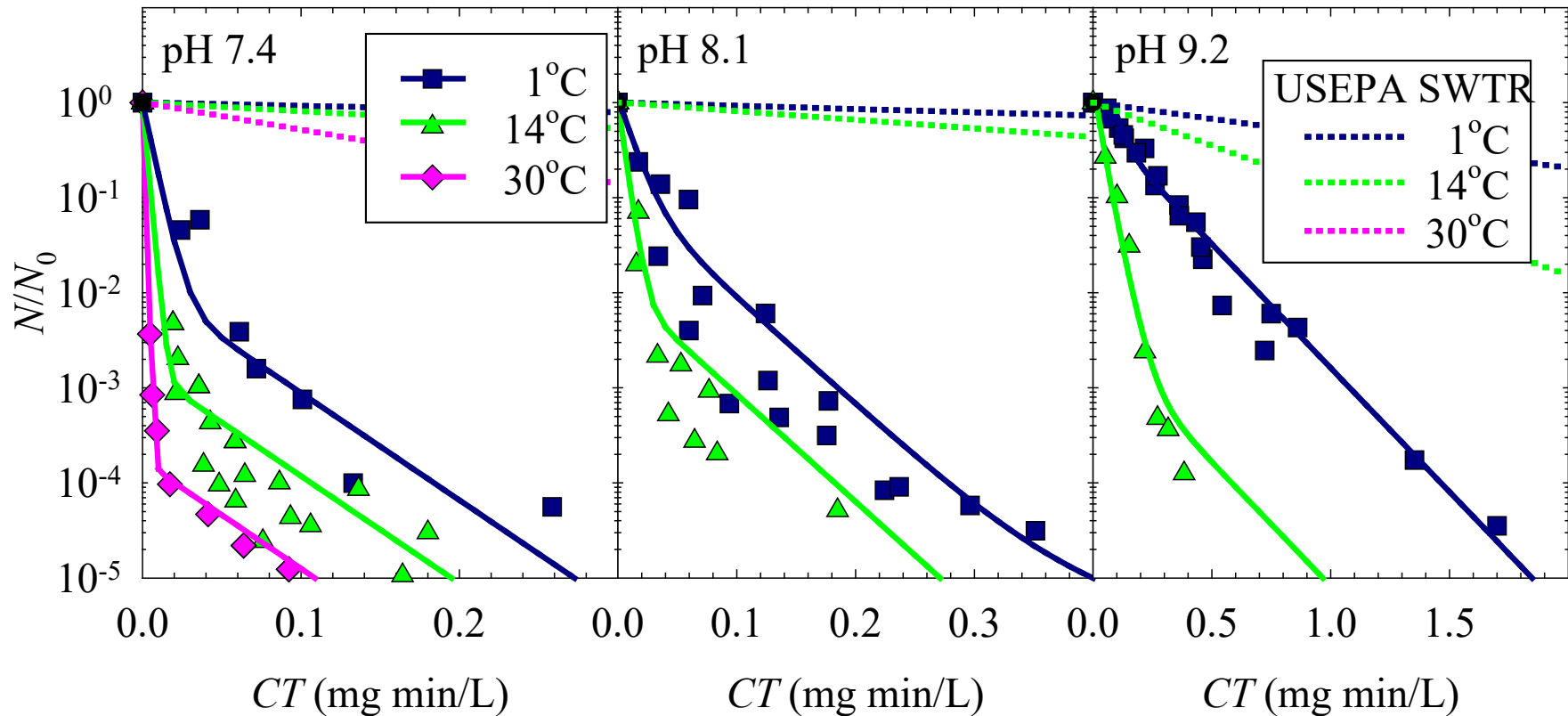


A.M. Gall, J.L. Shisler, B.J. Marinas, *Environ. Sci. Technol. Lett.*, 2016, 3, 185-189



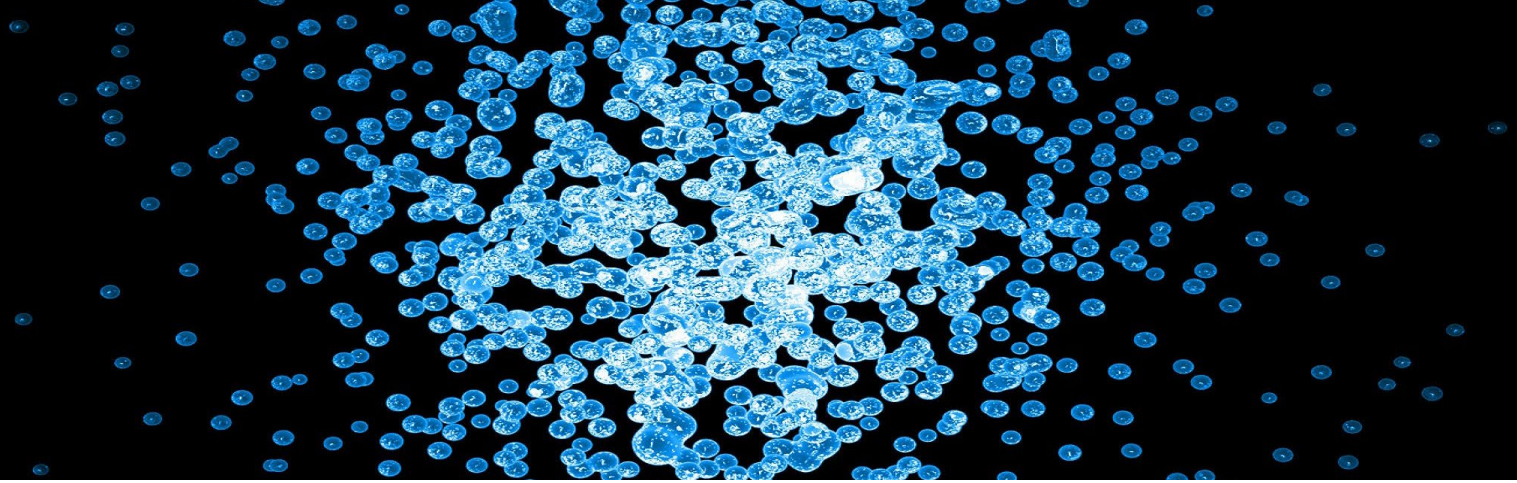
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Inactivation of Human Adenovirus 2 with Free Chlorine



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INACTIVATION MECHANISMS: REPLICATION CYCLE STEP INHIBITION

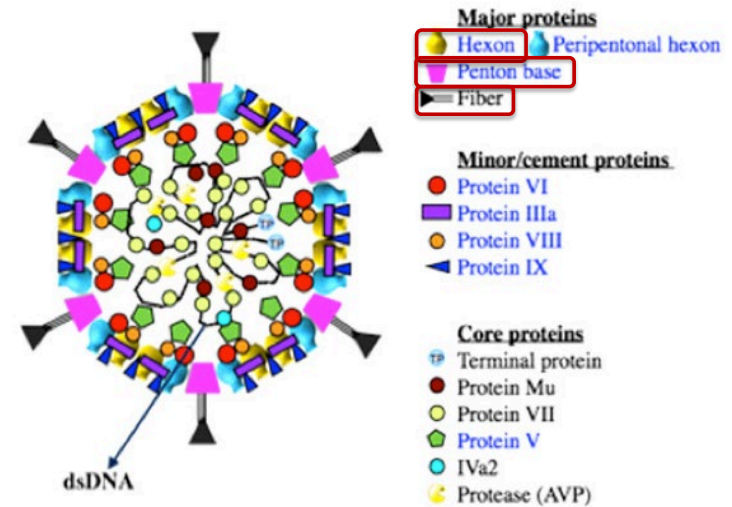
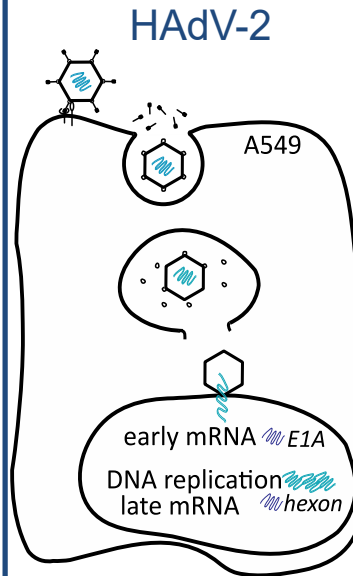


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HAdV-2 Replication Cycle

Selected HAdV-2 (90 nm, ~36,000 bp)
Replication cycle events:

- ▣ Attachment (fiber to CAR) – 0 h p.i.
- ▣ Internalization with endosome (penton base to integrin) – 15 min p.i.
- ▣ Virion uncoating and endosome rupture
- ▣ DNA release into nucleus
- ▣ Early mRNA (E1A) synthesis – 1-2 h p.i.
- ▣ DNA replication and late mRNA (hexon) synthesis – 5-8 h p.i.
- ▣ Cell lysis – 48 h p.i.



(Reddy and Nemerow, 2014)

Location of selected HAdV-2 DNA amplicons

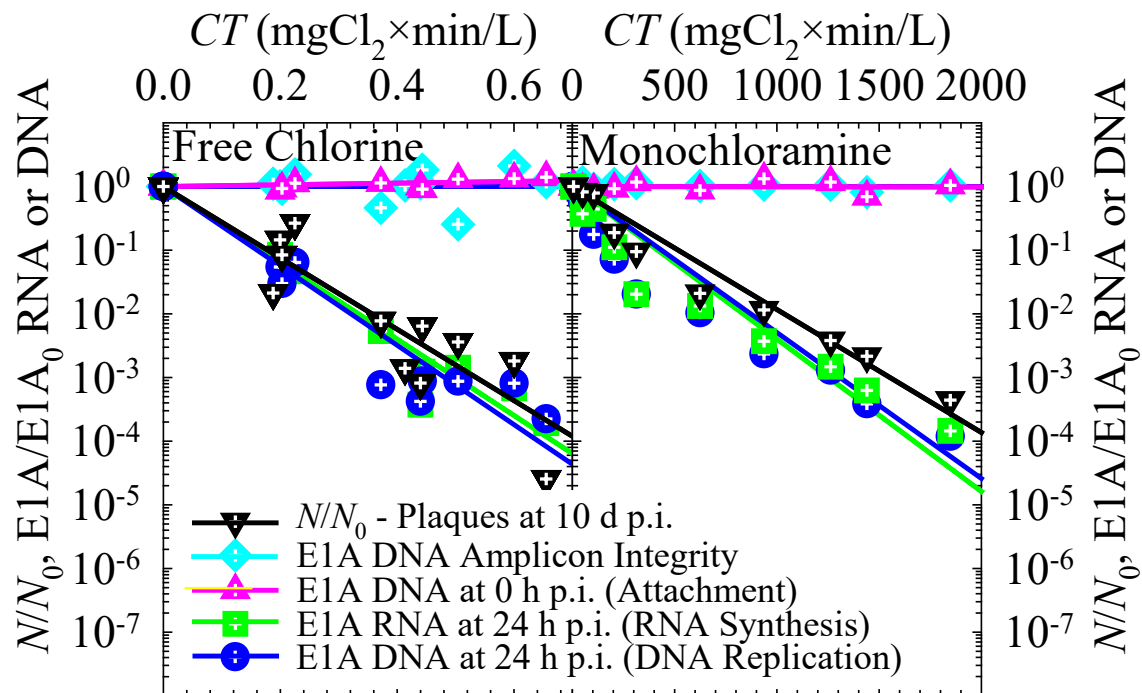


DNA: Quantitative Polymerase Chain Reaction (qPCR)
mRNA: Reverse Transcriptase qPCR (RT-qPCR)



Inactivation Human Adenovirus 2 with free chlorine (pH 9.2, 15°C) and monochloramine (pH 8, 15°C) compared to relative quantity of early (E1A) and late (hexon) DNA and RNA

- Adenovirus inactivation at levels up to 99.99% did not result in loss of amplicon integrity or ability to attach
- However, synthesis of viral DNA and early E1A and late hexon gene transcription were inhibited



A.M. Gall, J.L. Shisler, B.J. Mariñas, *Environ. Sci. Technol.*, 2015, 49, 4584-4590

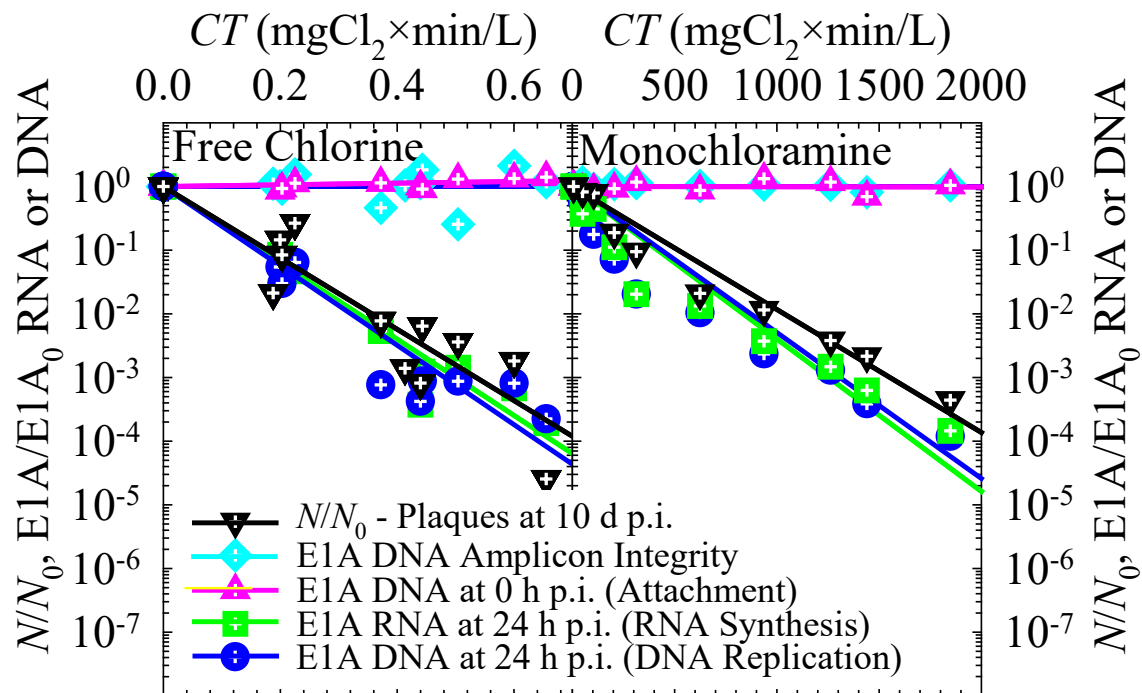
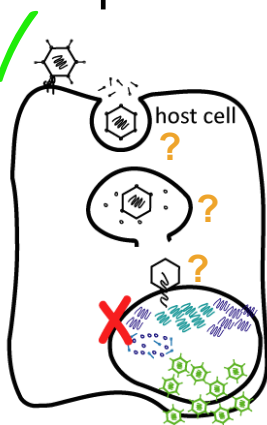
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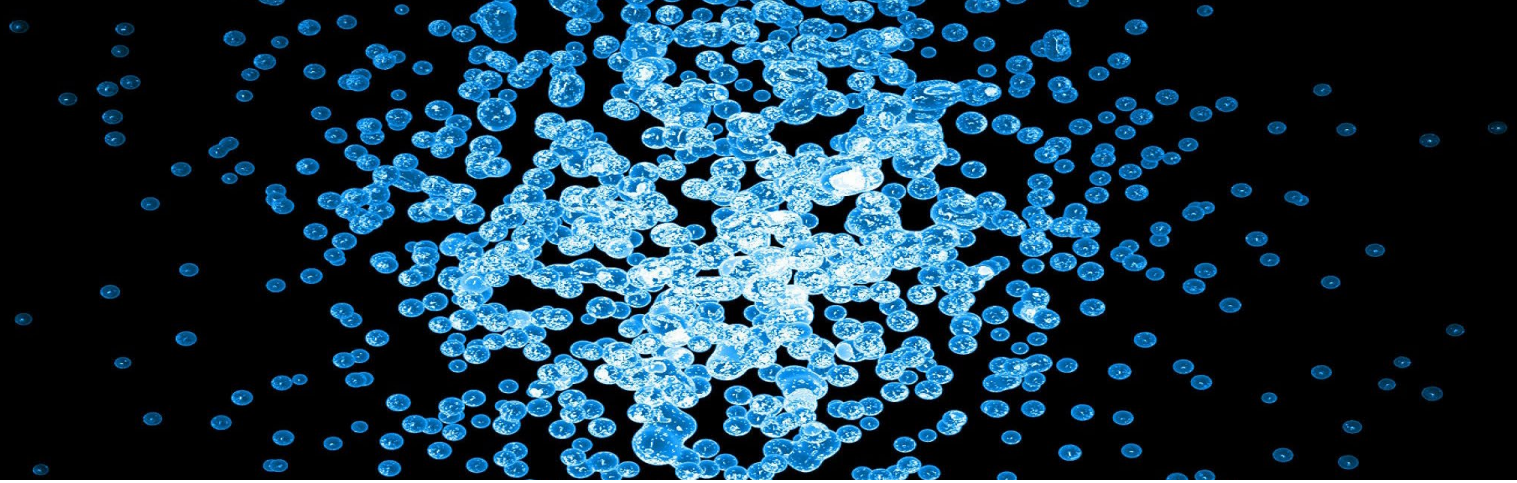
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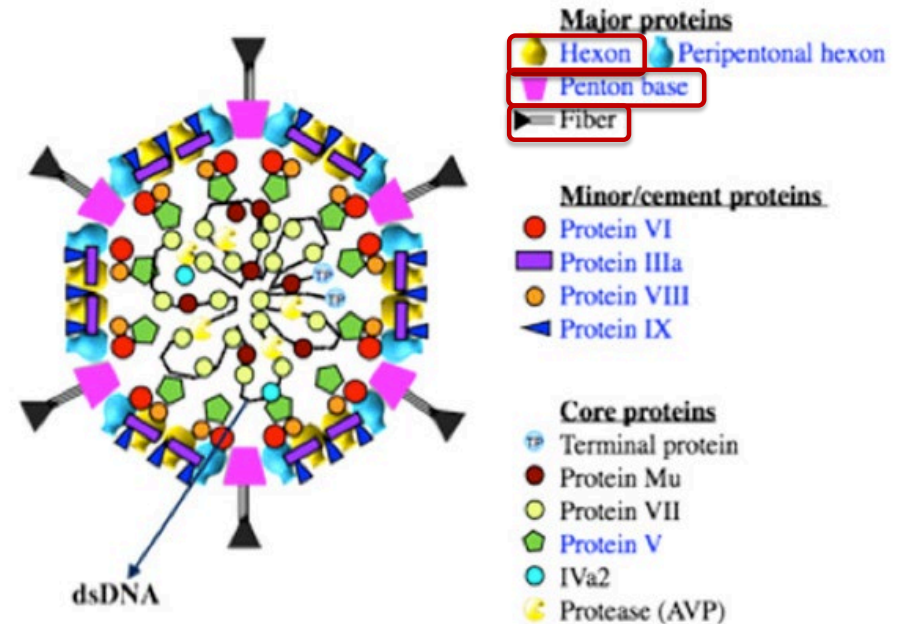
INACTIVATION MECHANISMS: CAPSID PROTEIN TRANSFORMATION



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Modifications of Human Adenovirus 2 Capsid Proteins with Free Chlorine

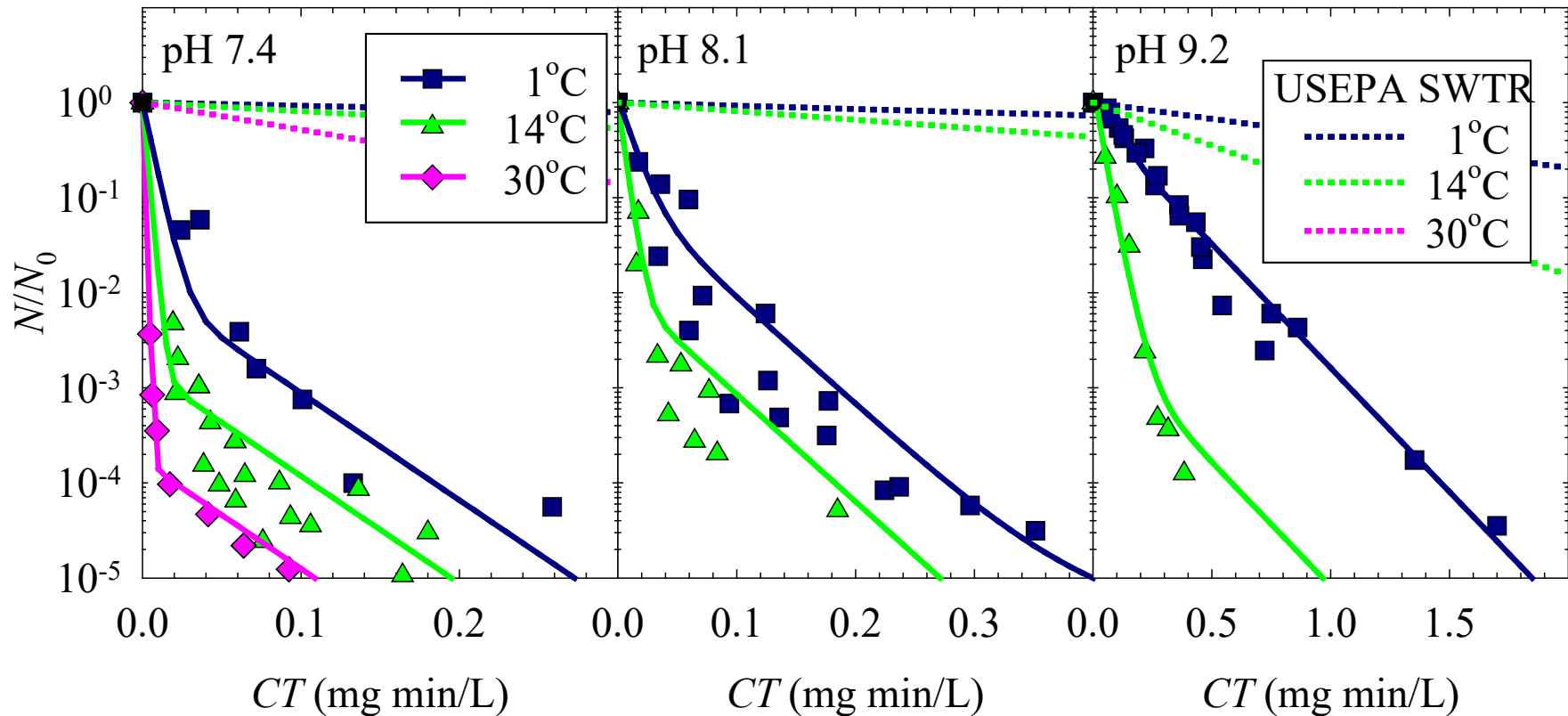
- Proteins targeted were: fiber, penton base, hexon
- The genes of the adenovirus fiber, penton base, and hexon were expressed in gene-modified *E. coli*
- 1 μM of each protein monomer (with 571, 582, 967 residues) was reacted with 100 μM free chlorine for 5 min ($CT \approx 3.6$ mg min/L, pH 8, 22°C) before quenching with excess sodium thiosulfate
- Proteins digested (Trypsin) and residues analyzed by LC-MS/MS



(Reddy and Nemerow, 2014)



Inactivation of Human Adenovirus 2 with Free Chlorine

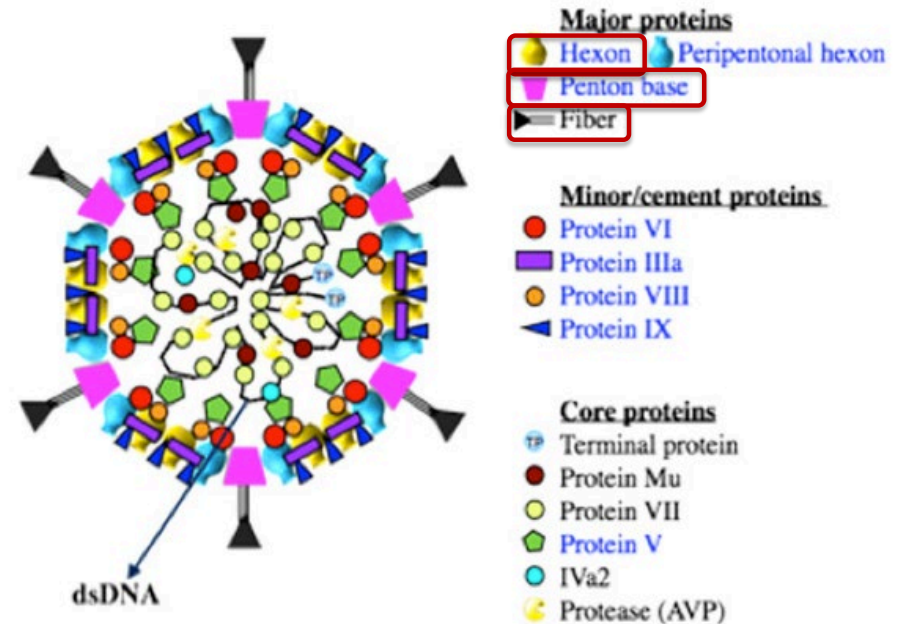


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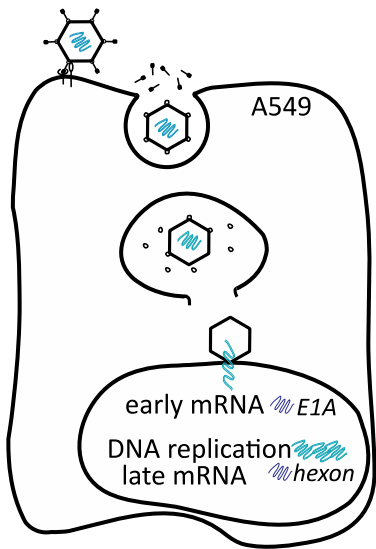


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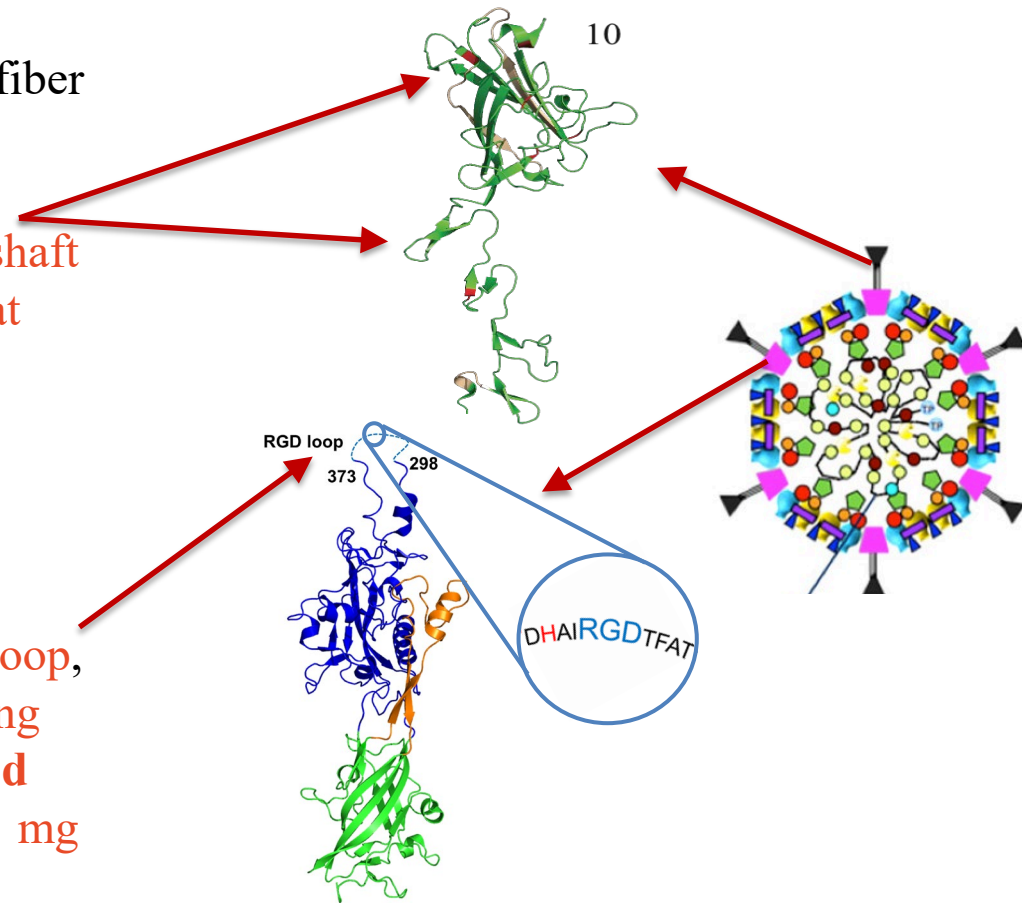
Graphical depictions of the HAdV-2 fiber, penton based, and hexon (bottom) fiber (expressed in *E. coli*) modifications by free chlorine exposure

Met≈Cys>His>Trp>Tyr>Asn≈Gln



• Ribbon diagram of fiber tail, shaft, and head domains: **CAR D1 binding & flexible shaft regions preserved at $CT=3.6$ mg min/L;**

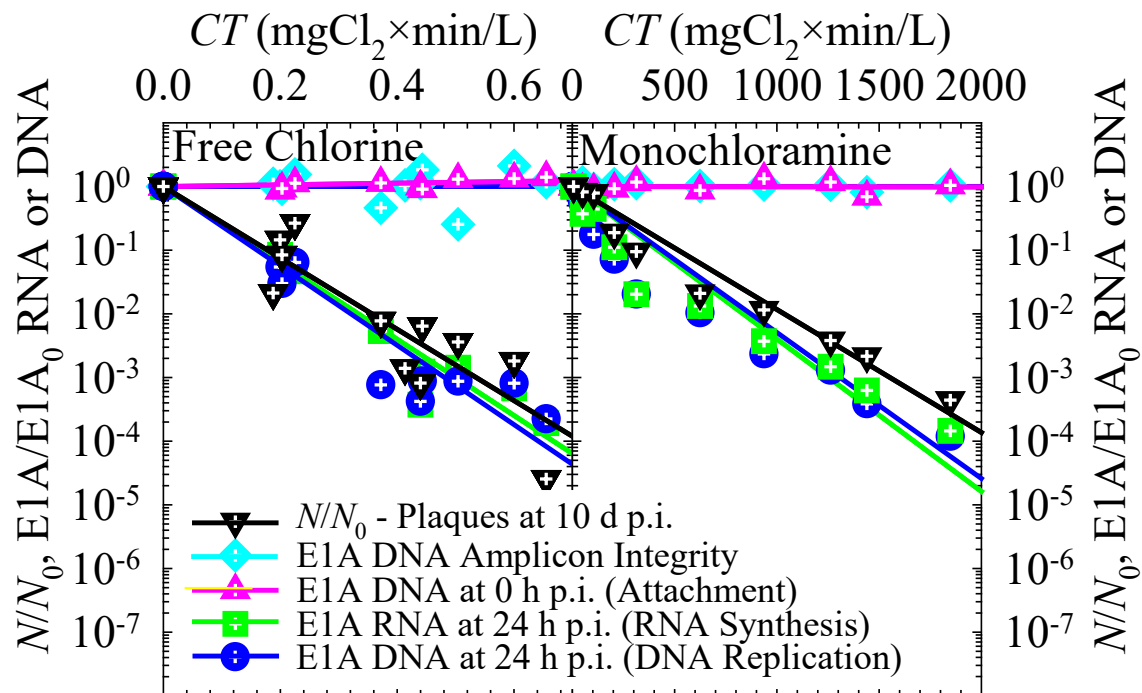
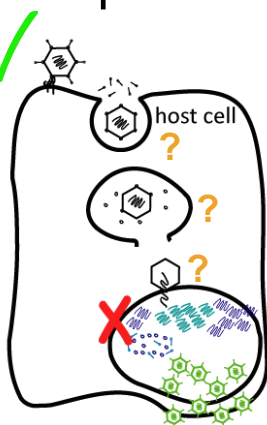
• Ribbon diagram of penton base: **RGD loop, RGD integrin binding regions transformed (His337) at $CT=3.6$ mg min/L;**



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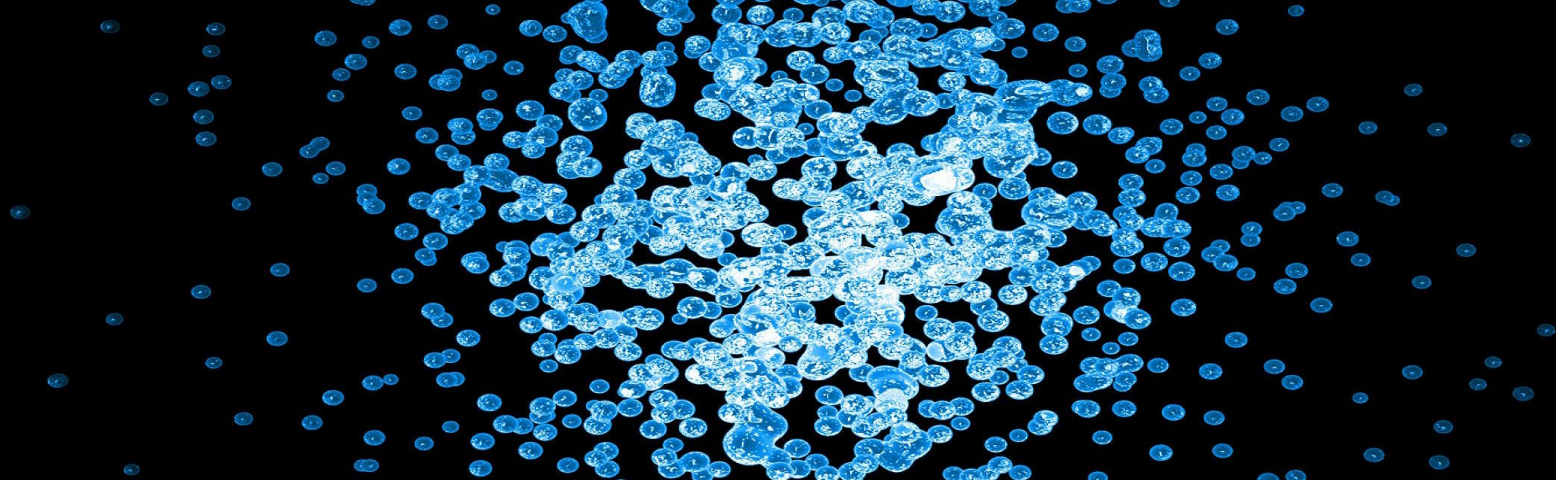
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INFECTIOUS VIRUS DETECTION: APTAMER-BASED SOLID STATE NANOPORE SENSOR DEVELOPMENT



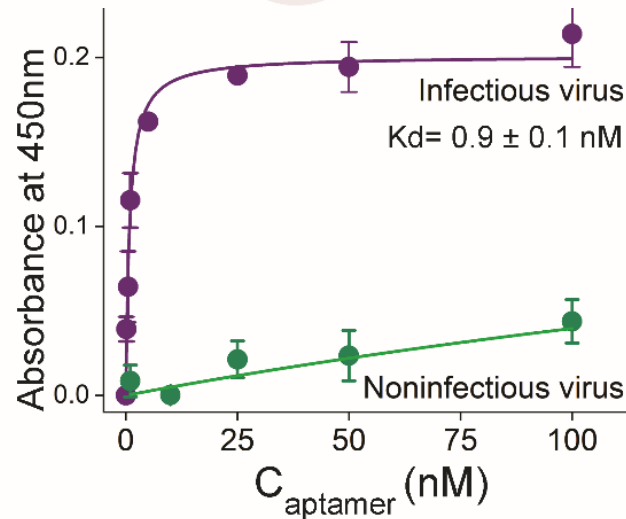
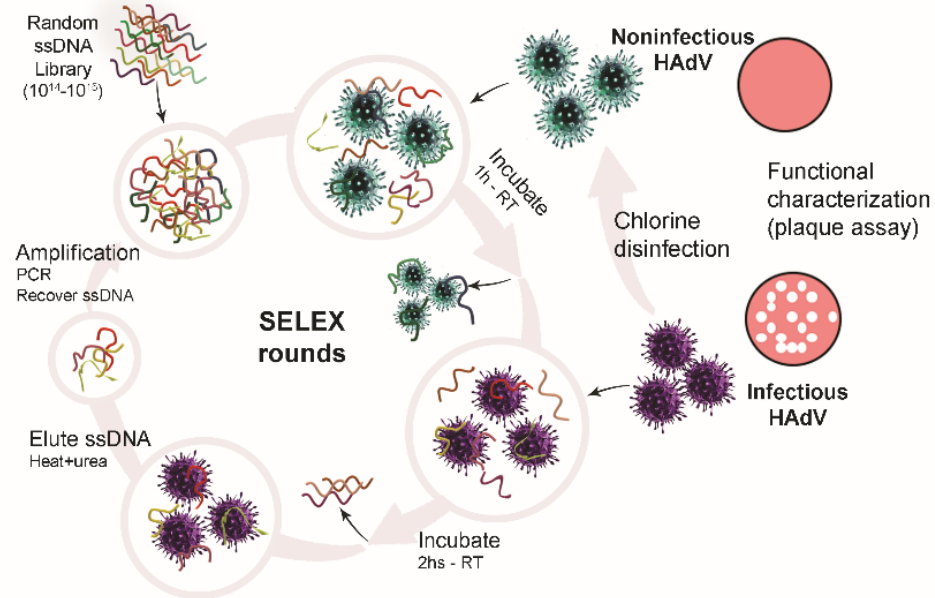
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In vitro selection of infectious HAdV-2-specific aptamer

• *in vitro* selection process for infectious HAdV-2 resulting in HAdV-Seq4 aptamer: positive and counter selections steps (the latter using non-infectious viruses) added in each round to reach high specificity toward infectious virus.

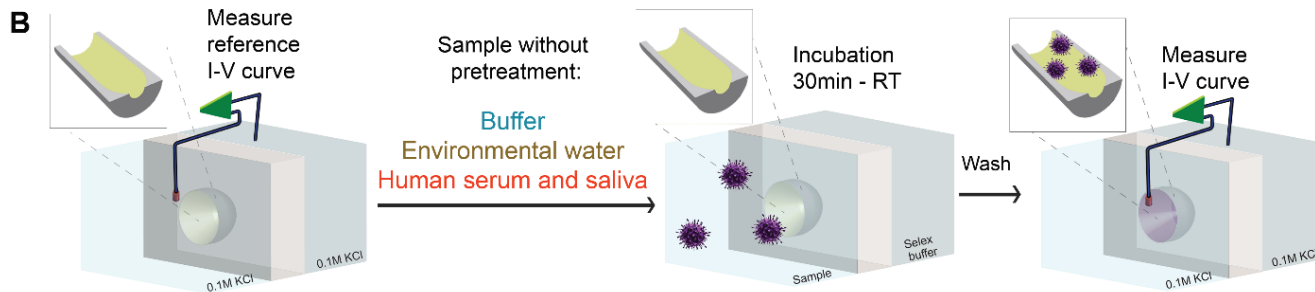
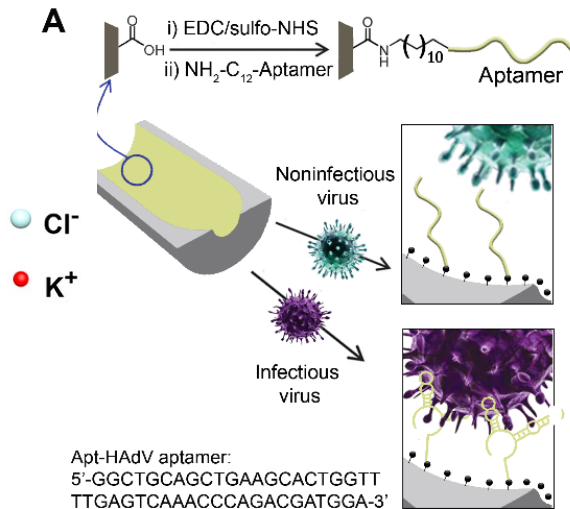
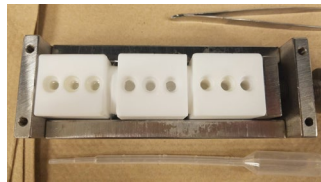
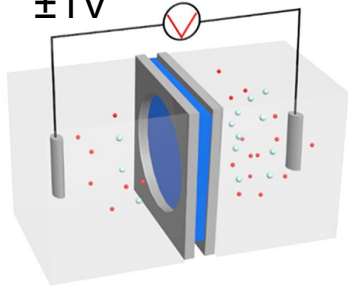
• Binding curves from affinity ELONA assays: dissociation constant ($K_d=0.9$ nM) of the HAdV-Seq4 aptamer for infectious HAdV-2 was more than 100 times higher than that for noninfectious HAdV-2.





Solid State Nanopore Sensor

- electrochemical cell: 0.1 M KCl, $\pm 1V$



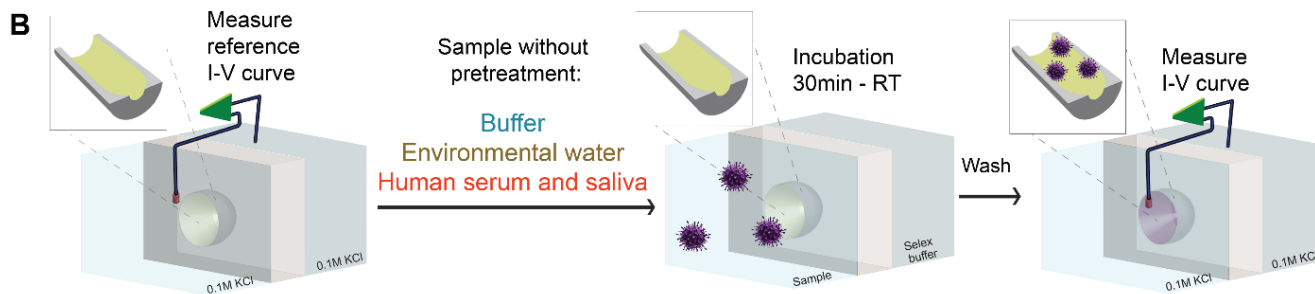
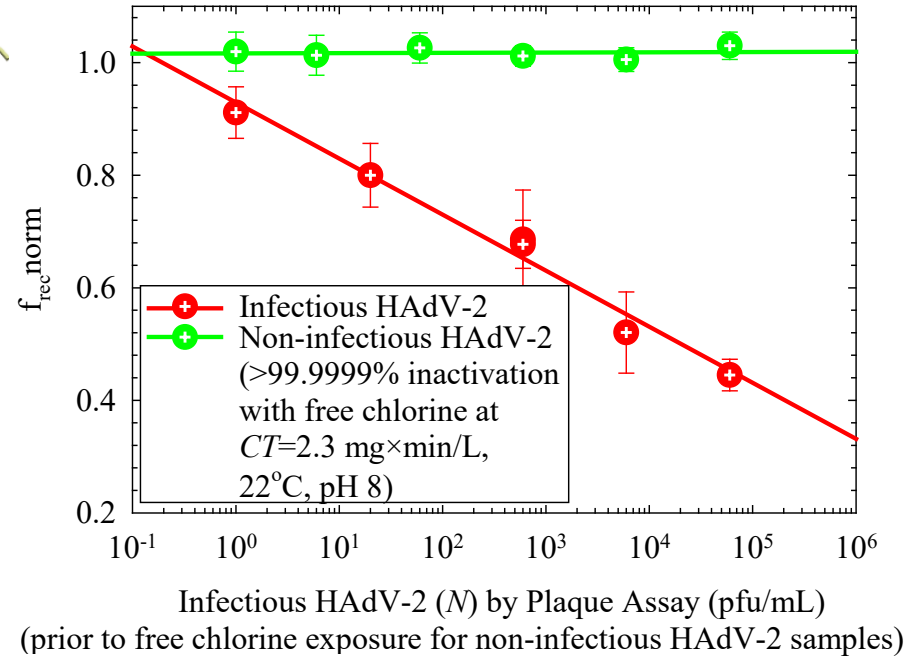
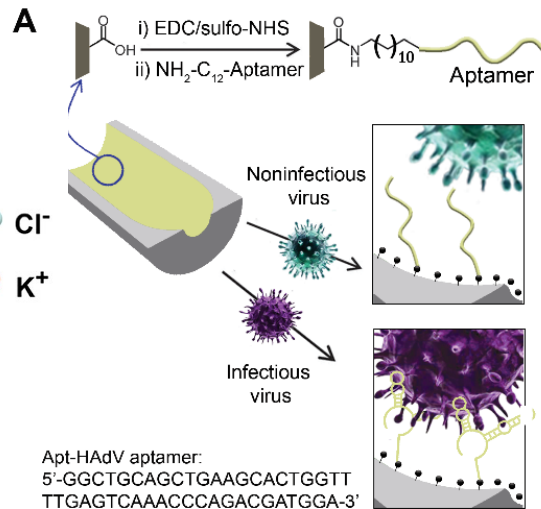
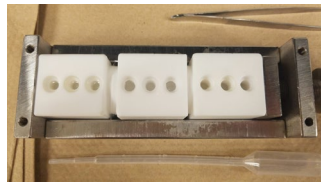
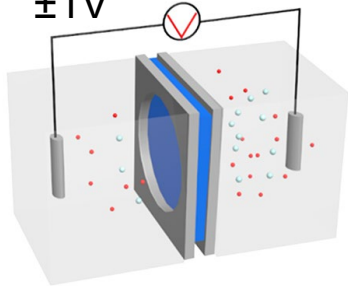
- The aptamer showed great selectivity for the infectious (active) HAdV
- The nanopore (900 \pm 100 nm \rightarrow 55 \pm 5 nm) amplified the signal several orders of magnitude





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A.S. Peinetti, R.J. Lake, W. Cong, L. Cooper, Y. Wu, Y. Ma, G.T. Pawel, M.E. Toimil Morales, C. Trautman, L. Rong, B.J. Marinas, O. Azzaroni, Yi Lu, *Sci. Adv.* (under revision)

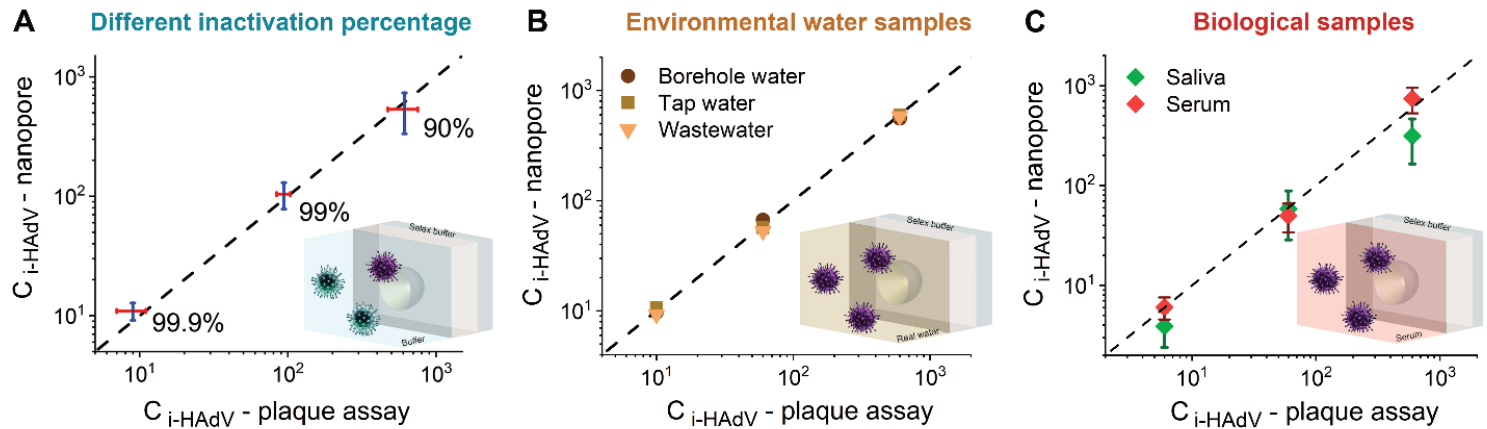


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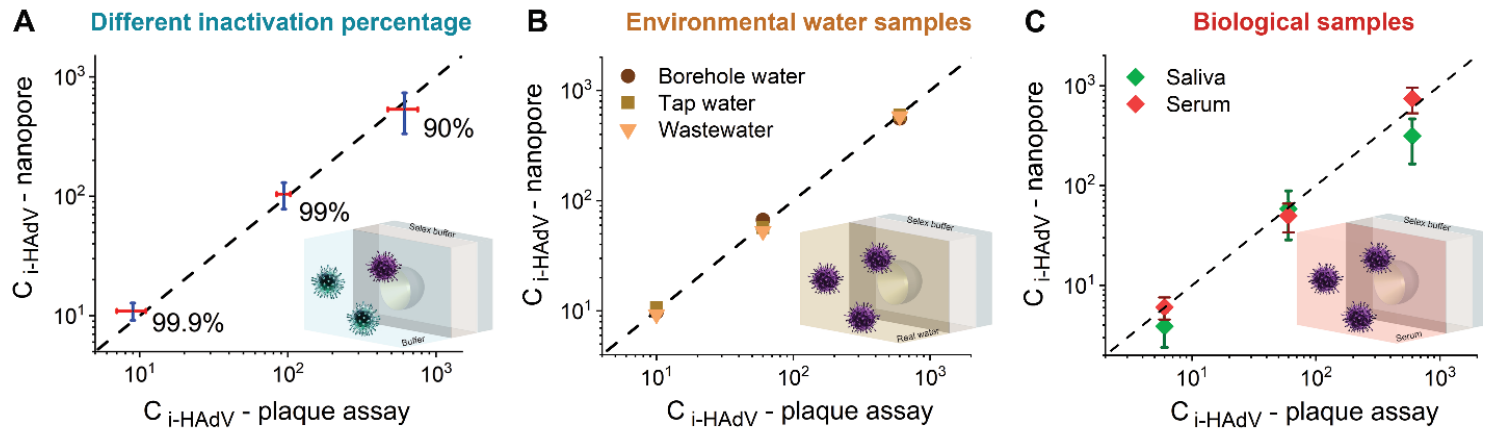
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- No major background interference in buffer, tap water, WW effluent, saliva, blood serum



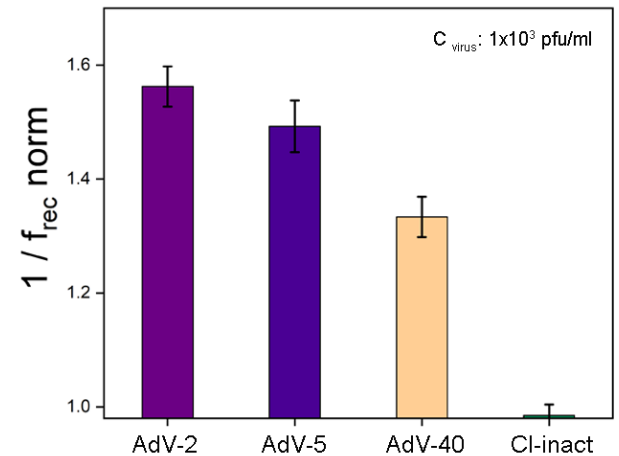
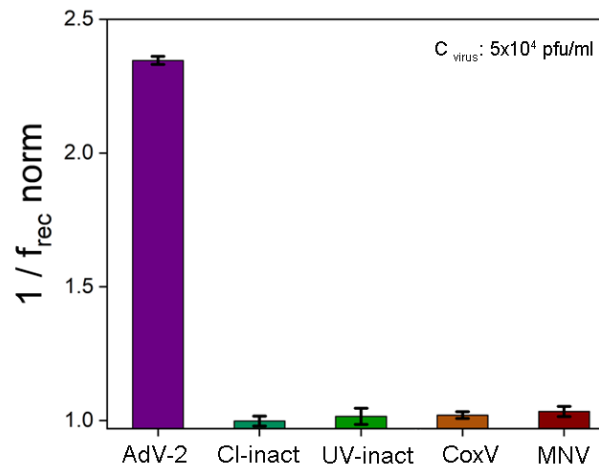


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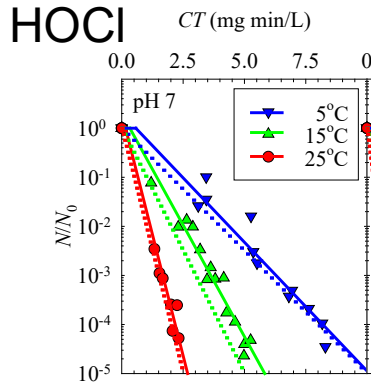


- Preliminary selectivity tests

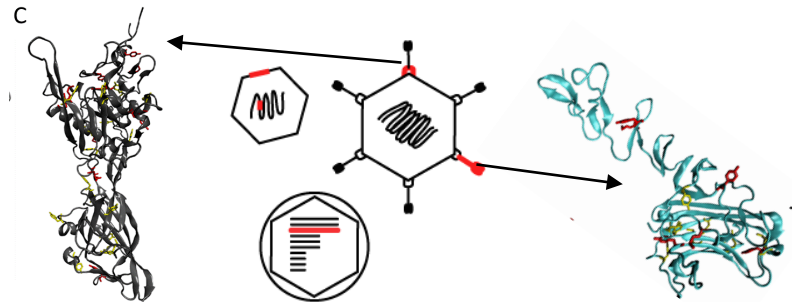


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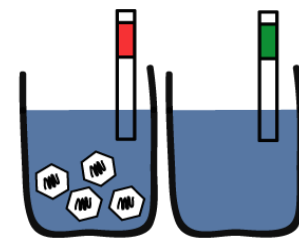
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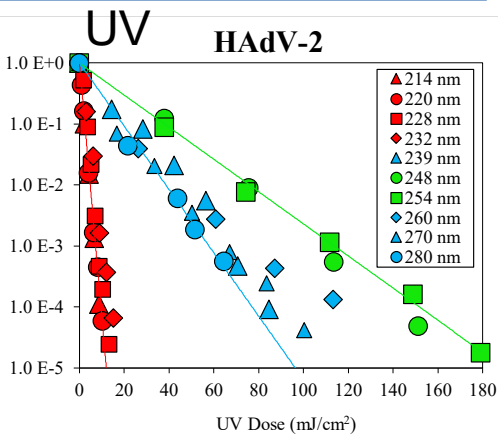
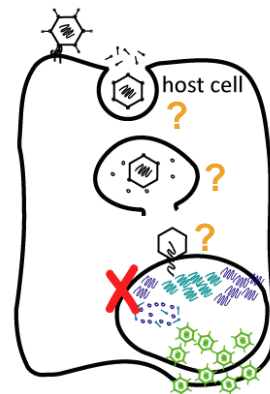


Real time detection of infectious viruses

Sensor development



Gall et al., *PLoS Pathog* 2015, 11 (6), e1004867



THANK YOU!

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